

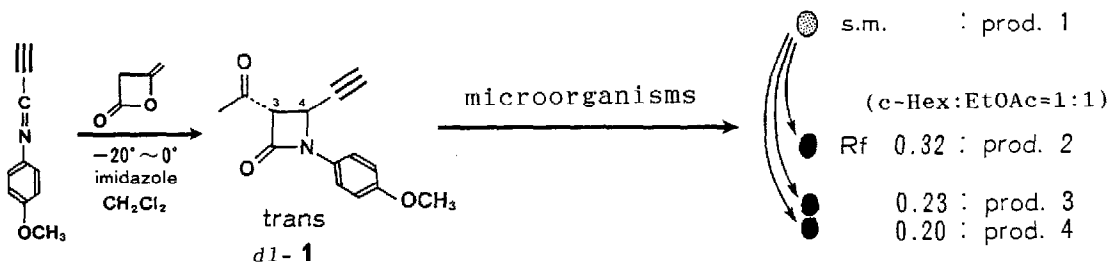
MICROBIAL TRANSFORMATION OF *dl* 3-ACETYL-AZETIDINONE DERIVATIVES¹⁾

Koichi Hirai* and Atsushi Naito

New Lead Res. Lab.* and Fermentation Res. Lab., Sankyo Co. Ltd.,
1-2-58 Hiromachi, Shinagawa-ku, Tokyo 140, Japan

Summary: Monocyclic β -lactam, *dl*-1 prepared by the diketene cycloaddition method was subjected to microbial transformation to give 3,4-trans-(R),(S)-OH mixture 2 and 3,4-cis-(R)-OH 3 and 3,4-cis-(S)-OH 4 depending on the microorganisms.

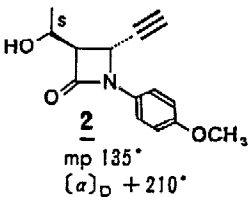
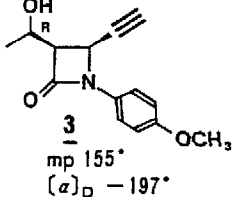
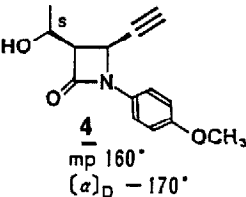
Current interest in the application of the microbial or enzymatic technique to the field of organic synthesis broaden the gate into the stereo- and regio-specific synthetic methodologies, particularly the enantio-specific syntheses of the chiral compounds²⁾. In the course of our study on the β -lactam chemistry the microbial transformation of the racemic β -lactam derivative such as 1 seemed to be attractive in the expectation for the specific reduction of the acetyl group on the azetidinone molecule. The starting material, *dl* 3,4-trans-1-(*p*-methoxyphenyl)-3-acetyl-4-ethynyl-2-azetidinone 1, mp 85°C was easily prepared by the method of Sumitomo group using diketene [2+2] cycloaddition³⁾: To a solution of propargyl aldehyde (4g) in benzene (50ml) was added *p*-anisidine (9g) and 4A-MS (4g) and the mixture was stirred for 30 min at 20°C. After filtration the solvent was removed *in vacuo* and the residue was dissolved in methylene chloride (60ml), and imidazole (4.3g) was added. The whole mixture was cooled to -20°C and added diketene (7.8g) slowly keeping the reaction temperature at -20°C ~ -10°C. The reaction temperature was raised to 20°C (~ 2 hr). Methylene chloride (40ml) was added and the solution was washed with aqueous dil. HCl and H₂O successively and dried over MgSO₄. After removal of the solvent *in vacuo* the desired compound was isolated by silica gel rapid chromatography



(c-Hex:EtOAc=3:1) to give 5.7 g (32%) of the 3-acetyl derivative 1. TLC Rf=0.5 (c-Hex:EtOAc=1:1). NMR (CDCl₃) δ : 2.35 (3H,s), 2.50 (1H,d,J=2 Hz), 3.75 (3H,s), 4.33 (1H,d,J=2 Hz), 4.91 (1H,t,J=2 Hz), 6.7-7.5 (4H,A₂B₂ type). IR (Nujol) cm⁻¹: 1760, 1720, 2100.

The starting material in hand we examined at first the microbial transformation of this *dl* trans-3-acetyl derivative 1, and found mainly three products; prod.2,3 and 4 on silica gel TLC as illustrated. The products distribution was classified to the combination of prod.2, 2+3, 2+3+4 and 2+4 according to the microorganisms employed (about 40 species) and cultural conditions.

The representative results are shown in the next Table.

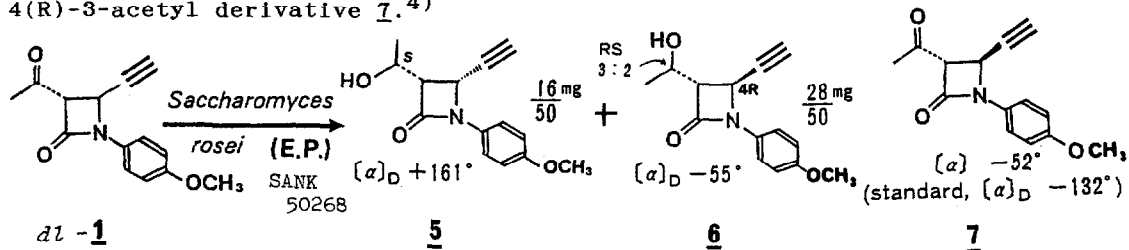
	prod. 2 3,4-trans R,S-OH mix.	prod. 3 3,4-cis R-OH	prod. 4 3,4-cis S-OH
S.P.: Stationary phase culture E.P.: Exponential phase culture	 <u>2</u> mp 135° [α] _D +210°	 <u>3</u> mp 155° [α] _D -197°	 <u>4</u> mp 160° [α] _D -170°
<i>Pichia terricola</i> SANK 51684 (S.P.)	$\frac{49 \text{ mg}}{100}$ [α] _D +184° S:R=3:1		$\frac{14 \text{ mg}}{100}$ [α] _D -144° ee 85%
<i>Saccharomyces cerevisiae</i> SANK 50161 (E.P.)	$\frac{90 \text{ mg}}{115}$ [α] _D +85° S only ee 40%		
<i>Schizosaccharomyces pombe</i> SANK 57362 (E.P.)	$\frac{30 \text{ mg}}{100}$ [α] _D +189° S only ee 90%	$\frac{37 \text{ mg}}{100}$ [α] _D -180° ee 91%	
<i>Trichosporon penicillutum</i> BY-356 (S.P.)	$\frac{35 \text{ mg}}{90}$ [α] _D +144° S only ee 69%	$\frac{25 \text{ mg}}{90}$ [α] _D -197° ee 100%	
<i>Streptomyces cattleya</i> SANK 63876 (S.P.)	$\frac{21 \text{ mg}}{100}$ [α] _D +144° S:R=15:1		$\frac{15 \text{ mg}}{100}$ [α] _D -169° ee 100%

[α]_D means [α]_D²⁴ (c=1, CHCl₃). $\frac{B}{A}$ mg means that A is the mg weight which was feeded and B mg is the mg weight obtained after silica gel TLC.

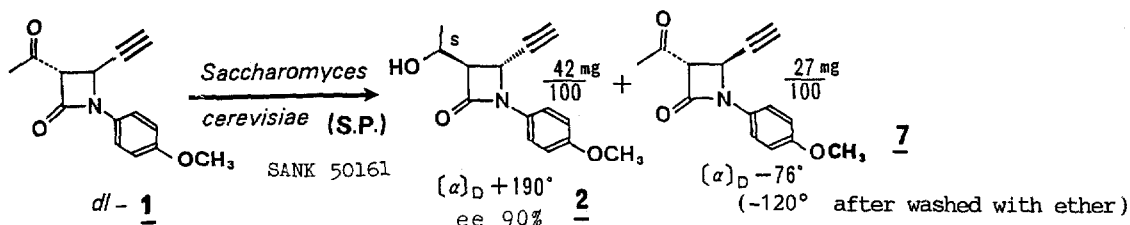
The structures of the each product were determined by the usual way: Product 2 was a mixture of 3,4-trans (R) and (S) hydroxyethyl azetidinones, but (S)-hydroxyethyl derivative 2 was predominant in every case. Recrystallization from ethylacetate and ether gave the pure prod.2, whose NMR is coincident with the structure 2 in which the coupling constant between C₃-H (δ 3.38,dd,J=2 and 4 Hz) and C₄-H (δ 4.60,t,J=2Hz) is 2 Hz. The absolute configuration was determined by the comparison of the chemically converted derivative, 3(R)-[1(S)-[(*tert*-butyldimethylsilyl)oxy]ethyl]-4(S)-[[[(phenylthio)carbonyl]methyl]azetidin-2-one [α]_D²⁴ -42°, with the standard enantiomer, [α]_D²⁴ +42° derived from penicillin⁵). Prod. 3 has

the coupling constant of 6 Hz (C_3 -H and C_4 -H) and C_3 -H appears at δ 3.50 as a triplet ($J=6$ Hz), and the prod.4 has the coupling constant of 6 Hz and C_3 -H appears at 3.43 as a doublet of doublet ($J=6$ and 9 Hz).

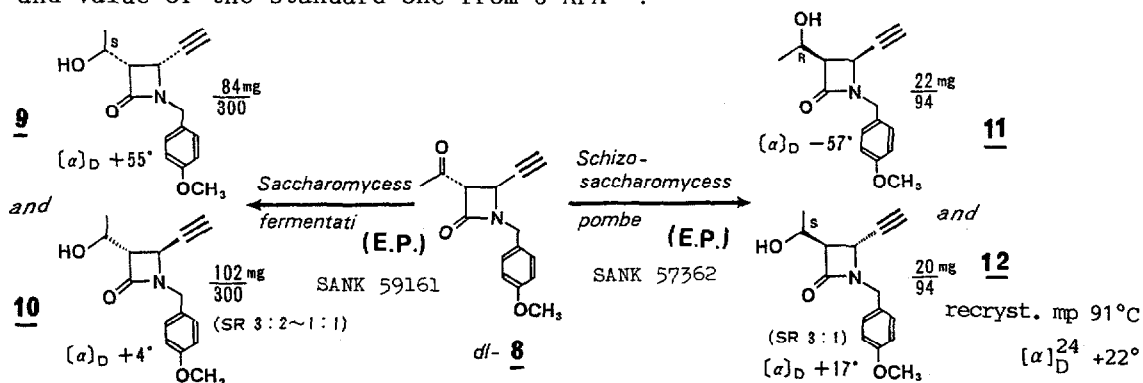
From these data and the other chemical criteria, and Dreiding model consideration we could determine the structure of the prod.3 and 4 as 3 and 4 which have (R) and (S) hydroxyethyl side chain with 3,4-cis configuration on the azetidinone molecule. Contrary to the above results in the case of *Saccharomyces rosei* two products, 5 and 6 were isolated, and one of which was proved to be 3,4-trans 4(R) derivative 6 (RS 3:2 from NMR) whose structure was confirmed by the comparison of the oxidation product of 6 with standard 4(R)-3-acetyl derivative 7.⁴⁾



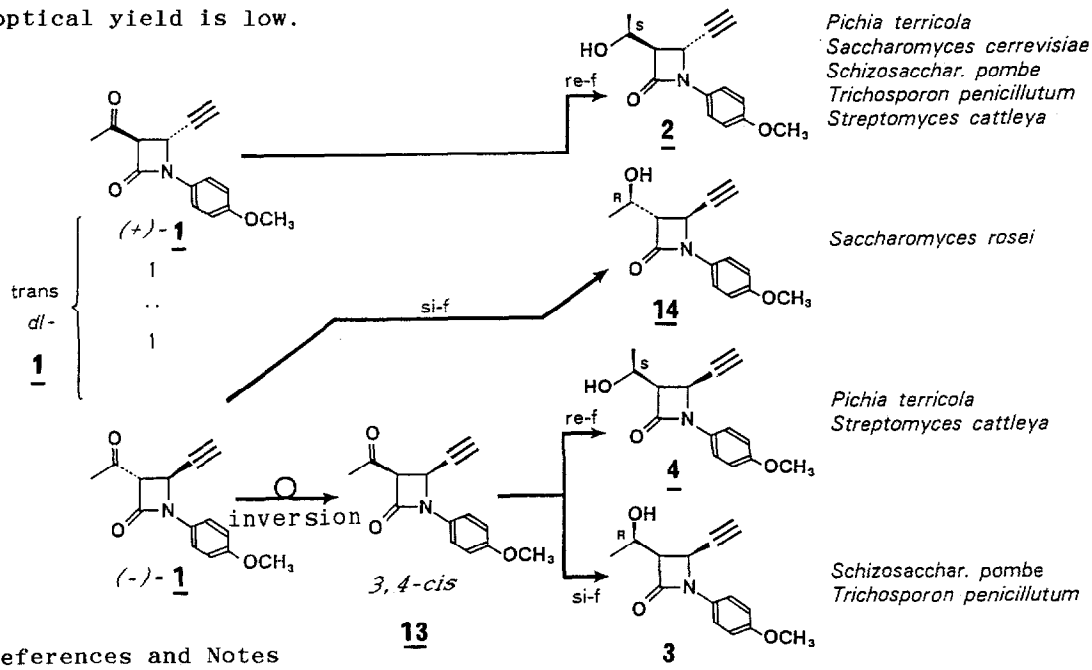
In the stationary phase (S.P.) culture we got (S)-alcohol 2 and the optically active 3-acetyl derivative 7 ($=(-)-1), which means exponential phase (E.P.) is more active than S.P. in our microbial reduction.$



In the case of *N*-anisyl derivative (*dl*-8) the parallel results were obtained, and one of the products 12 was recrystallized (mp 91°C , $[\alpha]_D^{24} +22^\circ$) and silylated to give silyl derivative $[\alpha]_D^{24} +17.5^\circ$, which is opposite signal and value of the standard one from 6-APA⁵⁾.

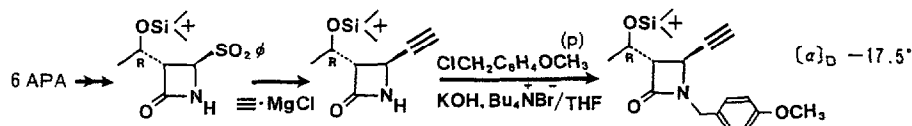


The summary of the microbial transformation products of the *dl* monocyclic β -lactam **1** is shown below. The acetyl group of the one enantiomer (+)-**1** was directly reduced to the (S)-hydroxyethyl product **2**. In the other enantiomer (-)-**1** the acetyl group was first converted to 3,4-cis intermediate **13** which was reduced (S)- or (R)-hydroxyethyl derivative **4** or **3** depending on the microorganisms. *Saccharomyces rosei* is different from other microorganisms so far examined because of the production of 3,4-trans with 3-(S), 4-(S) configuration, but the ratio of the R-hydroxyethyl to S-isomer is 3:2 and the optical yield is low.



References and Notes

- 1) A part of this work was presented at 106th Ann. Meeting Pharm. Soc. Japan 1986 and 16th IUPAC Int. Symp. Chem. Natural Prod. Kyoto, 1988.
- 2) 5th IUPAC Symp. Org. Syn. Freiburg, 1984. K. Mori and S. Senda, *Tetrahedron* **41**, 541 (1985). J.B. Jones, *Tetrahedron* **42**, 3351 (1986).
- 3) M. Sunagawa, K. Goda, M. Enomoto and A. Sasaki, *Abst. 9th Int. Cong. Heterocyclic Chem.*, p484 (1983, Tokyo). T. Kawabata, Y. Kimura, Y. Ito, S. Terashima, A. Sasaki and M. Sunagawa, *Tetra. Lett.* **27** 6241 (1986)
- 4) Obtained by PCC oxidation of prod. **3**.
- 5) From 6-APA by the following procedures the standard (-) silyl protected azetidinone derivative, $[\alpha]_D^{24} -17.5^\circ$ was obtained.
cf. T. Kobayashi, N. Ishida and T. Hiraoka, *Chem. Commun.*, 737 (1980).
H. Maruyama and T. Hiraoka, *J. Org. Chem.*, **51**, 399 (1986).



(Received in Japan 26 September 1988; accepted 23 December 1988)